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24.1 Summary

24.1.1 Topiramate and quetiapine are extracted from biological samples by making the samples basic with saturated borate buffer and extracting with toluene/hexane/isoamyl alcohol (THIA). An aliquot of the extract is analyzed by high performance liquid chromatography-electrospray ionization mass spectrometry (LC-ESI-MS).

24.2 Specimen Requirements

24.2.1 2 mL blood, fluid or tissue homogenate.

24.3 Reagents and Standards

- 24.3.1 Quetiapine, 1 mg/mL
- 24.3.2 Topiramate, 1 mg/mL
- 24.3.3 Mepivicaine, 1 mg/mL
- 24.3.4 Sodium tetraborate decahydrate
- 24.3.5 Hexane
- 24.3.6 Isoamyl alcohol
- 24.3.7 Methanol
- 24.3.8 Toluene
- 24.3.9 Ammonium acetate

24.4 Solutions, Internal Standard, Calibrators and Controls

- 24.4.1 10 mM Ammonium Acetate: Weight 0.389 g ammonium acetate. Transfer to 500 mL volumetric flask and QS to volume with dH₂O
- 24.4.2 Saturated borate buffer solution. Add sodium tetraborate decahydrate to dH_2O until no more dissolves after shaking vigorously.
- 24.4.3 Toluene:Hexane:Isoamyl Alcohol (THIA) (78:20:2, v:v:v) Mix 78 mL toluene, 20 mL hexane and 2 mL isoamyl alcohol.

24.4.4 Drug stock solutions:

- 24.4.4.1 If 1 mg/mL commercially prepared stock solutions are not available, prepare 1 mg/mL solutions from powders. Weigh 10 mg of the free drug, transfer to a 10 mL volumetric flask and QS to volume with methanol. Note: If using the salt form, determine the amount of the salt needed to equal 10 mg of the free drug, and weigh this amount. Stock solutions are stored capped in a refrigerator and are stable for 2 years.
- 24.4.5 Working standard solution for topiramate (0.1 mg/mL): Pipet 1 mL of the 1 mg/mL stock solution of topiramate into a 10 mL volumetric flask and QS to volume with methanol.

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- 24.4.6 Working standard solution for quetiapine (0.01 mg/mL): Pipet 100 μL of the 1 mg/mL stock solution of quetiapine into a 10 mL volumetric flask and QS to volume with methanol.
- 24.4.7 Working internal standard solution (0.1 mg/mL mepivicaine): Pipet 1 mL of the 1 mg/mL stock solution of mepivicaine into a 10 mL volumetric flask and QS to volume with dH₂O.
- 24.4.8 To prepare the calibration curve, pipet the following volumes of the 0.1 mg/mL topiramate working solution and 0.01 mg/mL quetiapine working solution into appropriately labeled 16 x 125 mm screw cap test tubes. Evaporate standards to dryness under nitrogen. Add 2 mL with blank blood to obtain the final concentrations listed below.

Amount of 0.1 mg/mL topiramate standard (μ L)	Final concentration of topiramate (mg/L)	Amount of 0.01 mg/mL quetiapine standard (μL)	Final concentration of quetiapine (mg/L)
1000	50	1000	5
400	20	400	2
200	10	200	1
100	5	100	0.5
40	2	40	0.2
20	1	20	0.1

24.4.9 Controls

- 10.1.1.1 Topiramate and Quetiapine Controls. Control may be from a external source or prepared in house using drugs from different manufacturers, lot numbers or prepared by a chemist different than the individual performing the extraction.
- 24.4.9.1 Negative control. Blood bank blood or equivalent determined not to contain topiramate, quetiapine or mepivicaine.

24.5 Apparatus

- 24.5.1 Test tubes, 16 x 125 mm, round bottom, borosilicate glass with Teflon caps
- 24.5.2 Test tubes, 16 x 114 mm, glass centrifuge, conical bottom
- 24.5.3 Centrifuge capable of 2000-3000 rpm
- 24.5.4 Nitrogen evaporator with heating block
- 24.5.5 Vortex mixer
- 24.5.6 GC autosampler vials with inserts
- 24.5.7 LC/MS: Agilent Model 1100 LC-MSD
 - 24.5.7.1 LCMS Instrument Conditions. The following instrument conditions may be modified to adjust or improve separation and sensitivity.

24.5.7.1.1 Elution Conditions

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24.5.7.1.1.1 Column: Agilent Hypersil BDS 125 mm X 3 mm, 3 µM particle size

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24.5.7.1.1.2 Column thermostat: 35° C

24.5.7.1.1.3 Solvent A: 55% 10 mM ammonium acetate

24.5.7.1.1.4 Solvent B: 45% methanol

24.5.7.1.1.5 Isocratic elution, stop time: 13.00 min

Time (min)	Solv. B	Flow
0.00	45	0.45
4.00	80	0.45
8.00	80	0.45
9.00	45	0.45

24.5.7.1.2 Spray Chamber

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24.5.7.1.2.1 Ionization Mode: Electrospray 24.5.7.1.2.2 Gas Temperature: 350° C 24.5.7.1.2.3 Drying Gas (N₂): 12.0 L/min 24.5.7.1.2.4 Nebulizer pressure: 30 psig 24.5.7.1.2.5 Vcap (Positive): 4000 V

24.5.7.1.3 Selected Ion Monitoring (quantitation ions)

24.5.7.1.3.1 Polarity: Positive 24.5.7.1.3.2 Injection volume: 2 μL

Time	Group Name	SIM Ion	Fragmentor	Gain	SIM	Actual
(min)				EMV	Resol.	Dwell
0.00	topiramate	264	150	1.5	Low	218
		282	150		218	
		340	150		218	
		<u>357</u>	150		218	
4.30	mepivicaine	98	150	0.5	Low	439
		<u>247</u>	150		439	
6.00	quetiapine	210	250	0.5	Low	218
		253	250		218	
		279	250		218	
		384	250		218	

24.6 Procedure

- 24.6.1 Label clean 16 x 125 mm screw cap tubes appropriately with calibrators, controls and case sample IDs.
- 24.6.2 Prepare calibrators and controls.
- 24.6.3 Add 2 mL case specimens to the appropriately labeled tubes.
- 24.6.4 Add 40 μ L 0.1 mg/mL mepivicaine internal standard working solution to each tube for a final concentration of 2mg/L.

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24.6.5 Add 2 mL saturated borate buffer and 6 mL extract solvent (78:20:2 THIA) to each tube.			

- 24.6.6 Cap and rotate tubes for 30 minutes.
- 24.6.7 Centrifuge at approx 2500 rpm for 15 minutes. Transfer organic upper layer (THIA) to appropriately labeled conical bottom test tubes.
- 24.6.8 Evaporate samples to dryness at approximately 50° C under nitrogen.
- 24.6.9 Reconstitute samples in 100 μL methanol. Vortex briefly. Transfer to GC autosampler vials for analysis by LCMS.

24.7 Calculation

24.7.1 Drug concentrations are calculated by linear regression analysis using the ChemStation software.

24.8 Quality Control and Reporting

24.8.1 See Toxicology Quality Guidelines

24.9 REFERENCES

- 24.9.1 M Contin, R Riva, F Albani and A Baruzzi. Simple and rapid liquid chromatographic-turbo ion spray mass spectrometric determination of topiramate in human plasma. J Chrom B 761: 133-137, 2001.
- 24.9.2 J Pearson and R Steiner, in-house development.